

Figure S1. Sorting nexin 9 (SNX9) specifically binds pSmad3 and not pSmad 1/5/8. Lysates from AKR-2B cells untreated (-) or stimulated (+) for 45 min with 5 ng/ml TGFβ or 10 ng/ml BMP4 were incubated with full length (FL) SNX9 immobilized on GST beads as in Figure 1A. Bound proteins were eluted and assessed by Western analysis for pSmad3 or pSmad1/5/8. Cell lysate reflects signal obtained from 10 μg total protein. Blots are representative of 3 separate experiments.

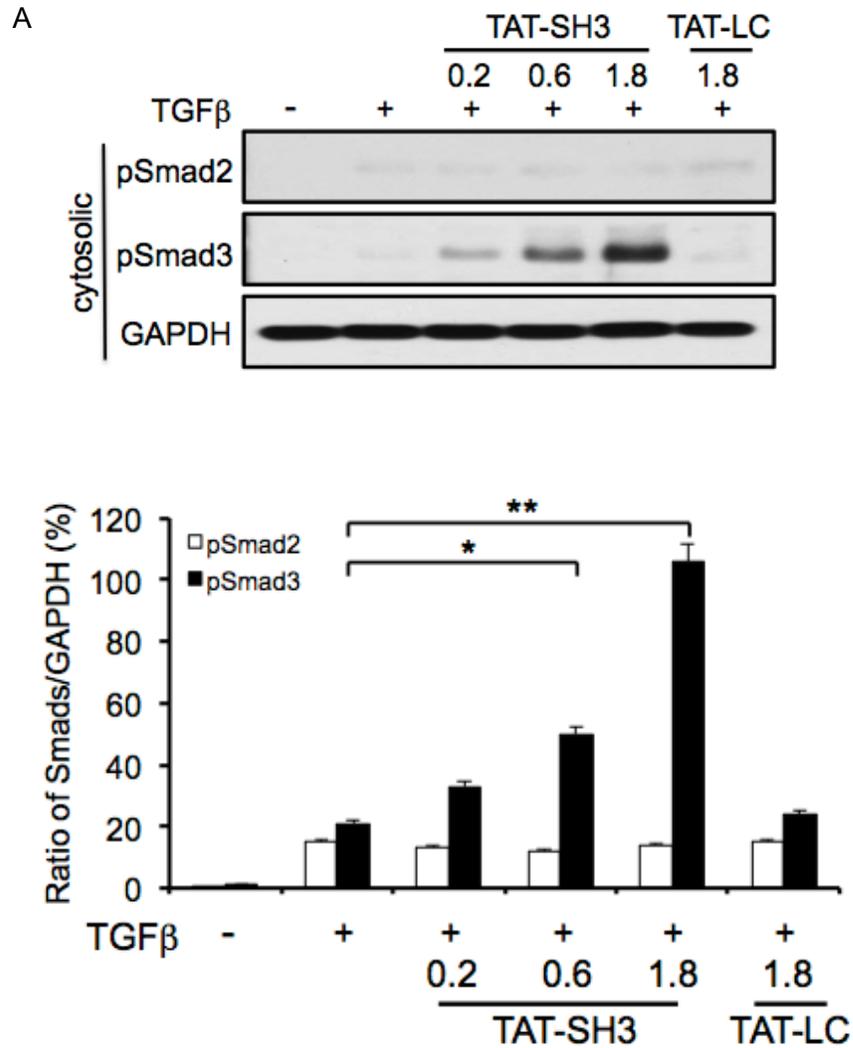
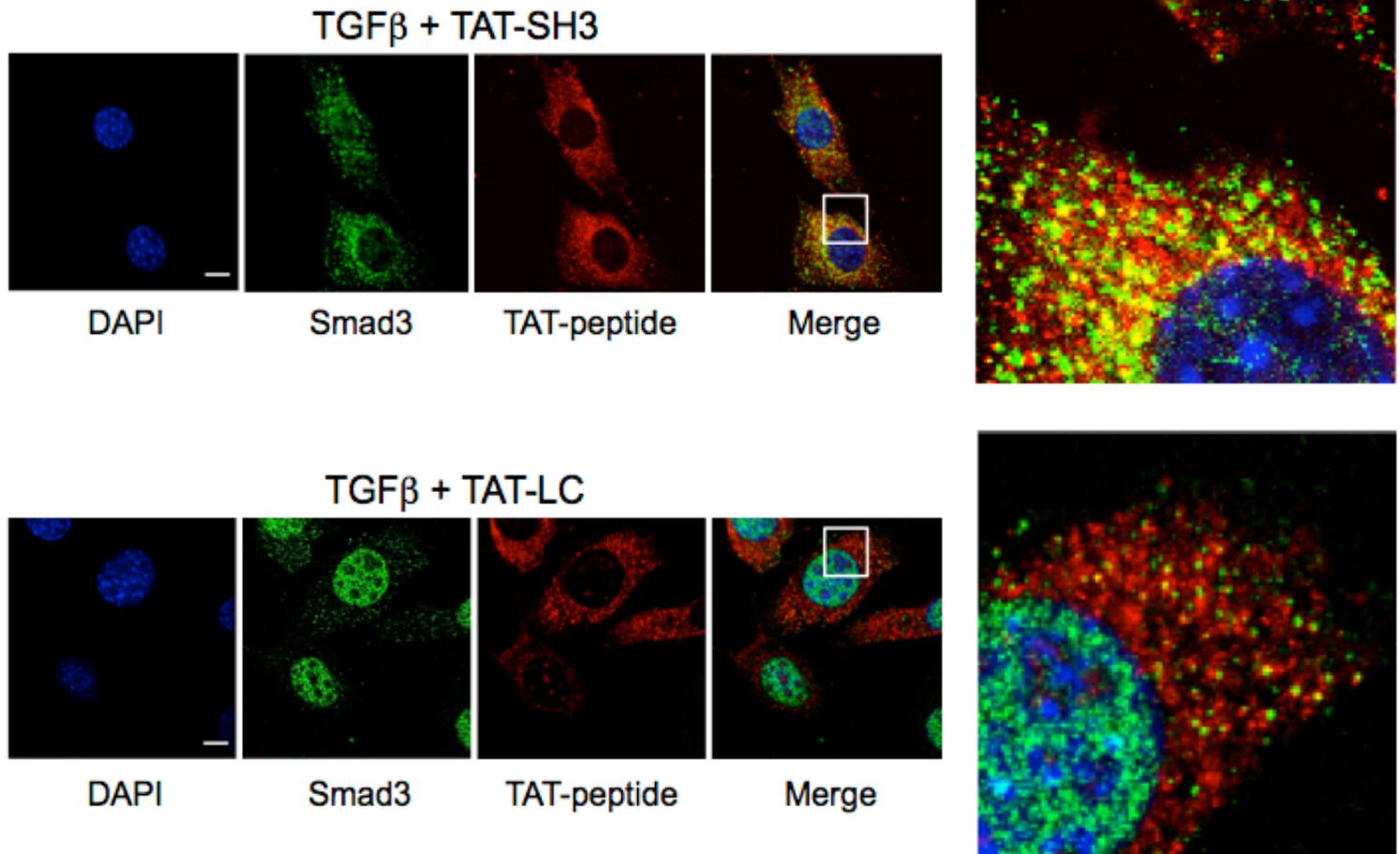


Figure S2. SH3 domain of sorting nexin 9 prevents nuclear pSmad3 import. (A) AKR-2B cells were transduced for 90 min with the indicated concentration (μM) of TAT fusion peptide to the SH3 or LC domains of SNX9 as in Figure 1B. Following washing and 1 hr TGF β (5 ng/ml) treatment cytosolic fractions were isolated and Western blotted for pSmad2, pSmad3, or GAPDH. Blots are representative of 3 separate experiments. (B) Quantitation of cytosolic pSmads was performed with Image J software and represents the mean \pm SEM of 3 experiments. Data was analyzed by one-way ANOVA followed by Dunnett's multiple comparisons test. * $P < 0.05$, ** $P < 0.005$.

A



B

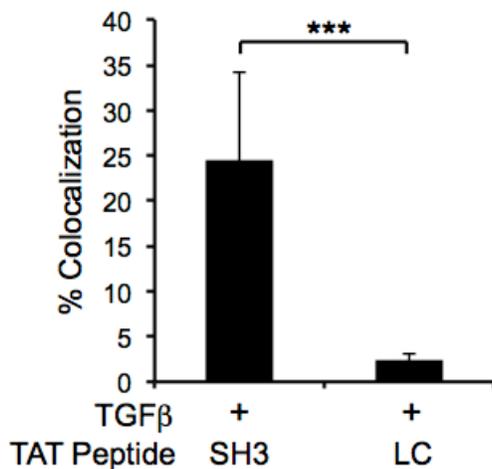


Figure S3. TAT-SH3 co-localizes with pSmad3. (A) AKR-2B cells were transduced for 90 min with TAT-SH3 or TAT-LC (1.8 μ M). Following washing and 45 min TGF β (5 ng/ml) treatment immunofluorescence for Smad3 or the HA-tagged TAT peptide was performed as described in Methods. Nuclei were stained with DAPI and right panels reflect higher magnification of boxed areas to visualize Smad3/TAT co-localization (yellow). (B). Quantitation of Smad3/TAT co-localization of 20 cells from each of 3 separate experiments using Image J software. Data was analyzed by 2-tailed Student's t-test. ***P<0.0005.

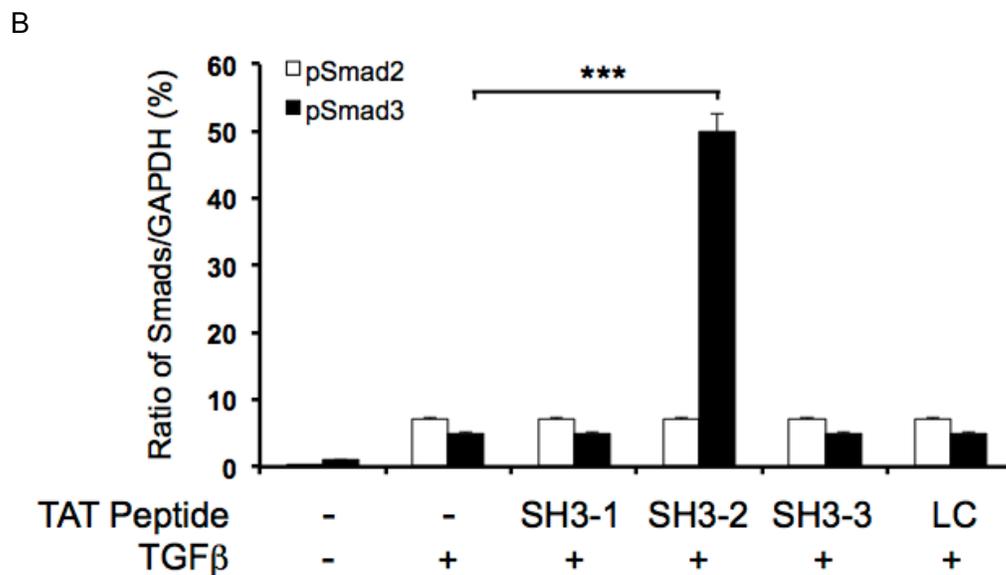
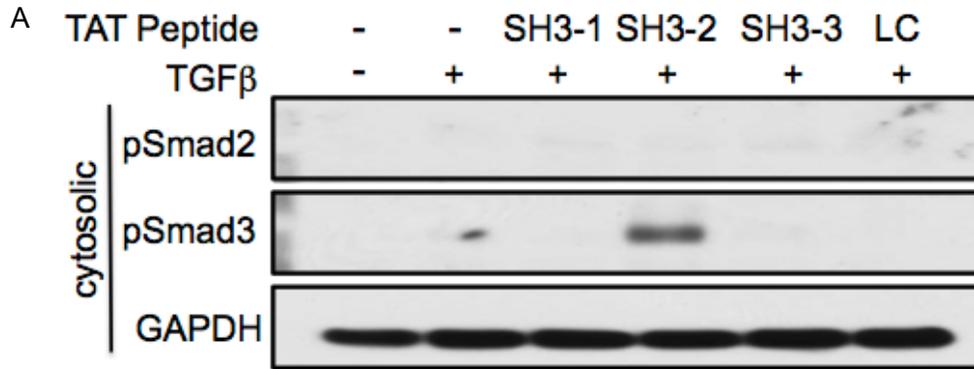


Figure S4. TAT-SH3-2 domain of sorting nexin 9 prevents nuclear pSmad3 import. (A) AKR-2B cells were transduced for 90 min with the indicated TAT peptide. Following washing and 1 hr TGFβ (5 ng/ml) treatment cytosolic fractions were isolated as in Figure S2 and Western blotted for pSmad2, pSmad3, or GAPDH. Blots are representative of 3 separate experiments. (B) Quantitation (mean +/- SEM) of cytosolic pSmad2 or pSmad3 from 3 experiments using Image J software. Data were analyzed by one-way ANOVA followed by Dunnett's multiple comparisons test. ***P<0.0005.

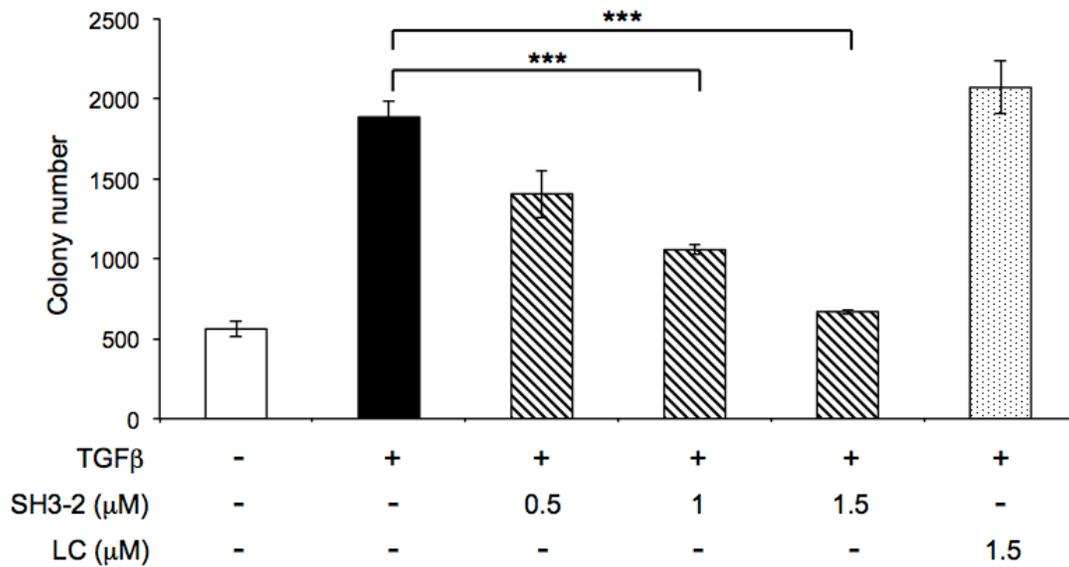


Figure S5. Dose-dependent inhibition of anchorage-independent growth by TAT-SH3-2. Soft agar colony formation was performed as described in Methods. Following 7 days growth in the presence (+) or absence (-) of TGFβ (20 ng/ml) or the indicated TAT peptide, colonies >50 mm were determined. Data reflect the mean +/- SEM of triplicate wells from 3 experiments. Data was analyzed by one-way ANOVA followed by Dunnett's multiple comparisons test. ***P<0.0005.

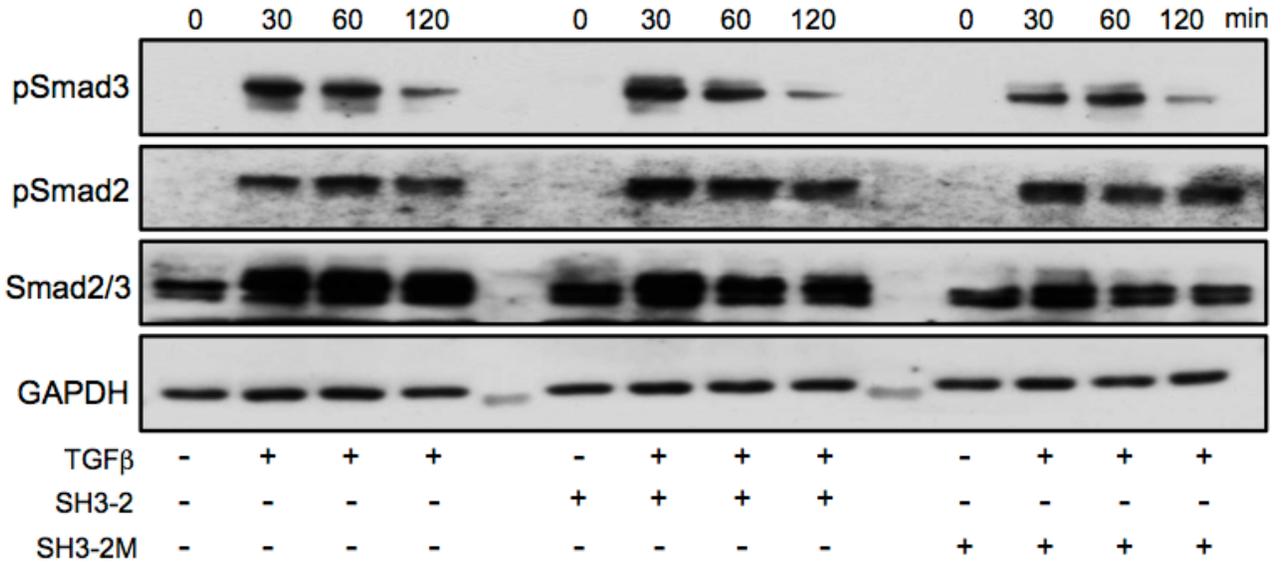
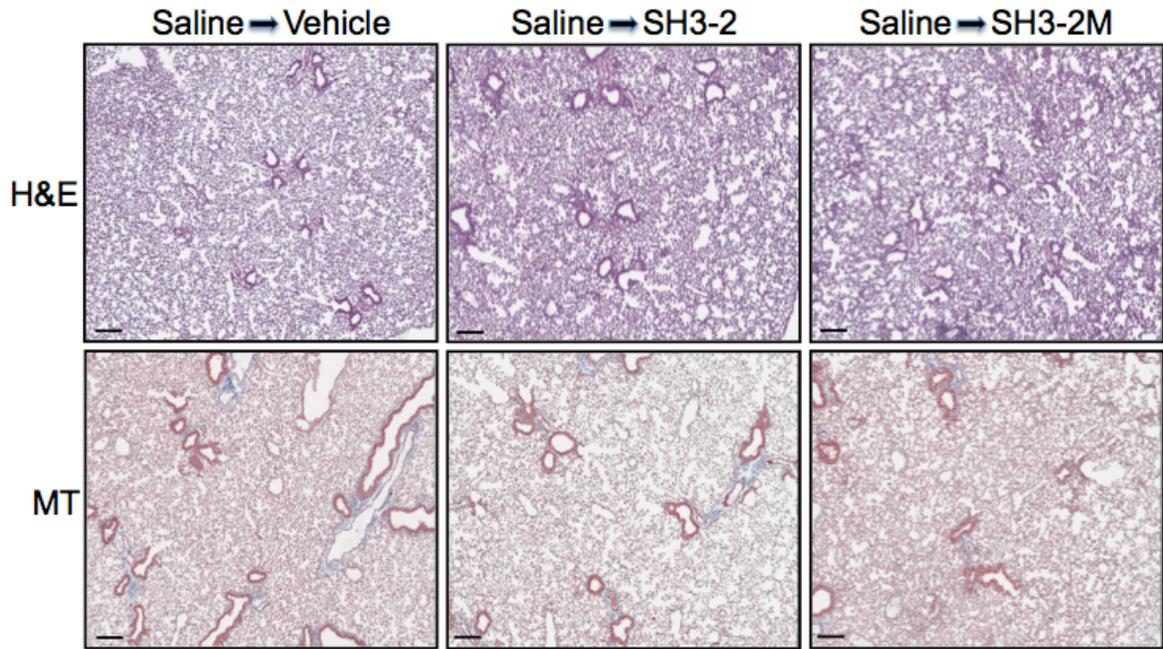


Figure S6. TAT-SH3-2 peptide does not affect Smad2 or Smad3 phosphorylation. AKR-2B cells were transduced for 90 min with 1.5 μ M TAT-SH3-2 or TAT-SH3-2M. After transduction, cells were treated (+) with TGF β (5 ng/ml) for the indicated times and processed by Western analysis for the indicated phosphorylated (p) or total protein. Band running slightly below GAPDH is the 34 kDa marker detected by the GAPDH sera. Blots are representative of 3 separate experiments.

A



B

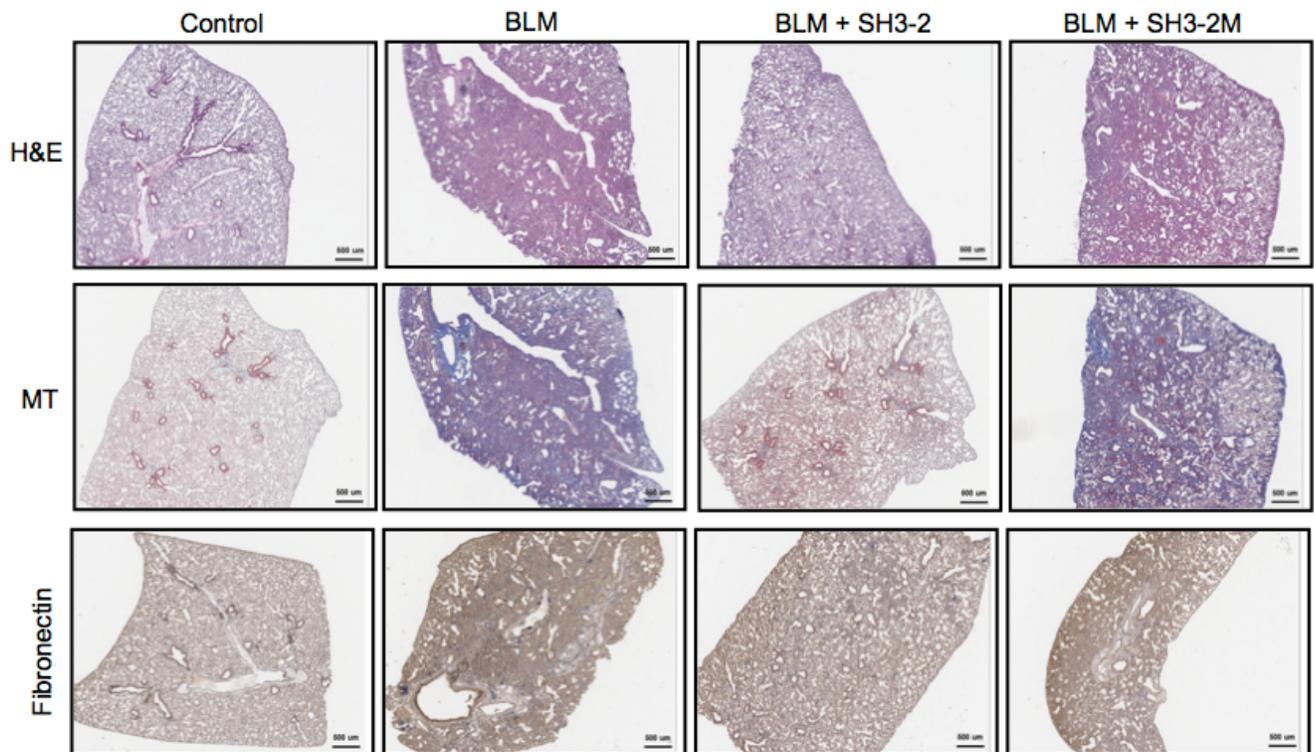
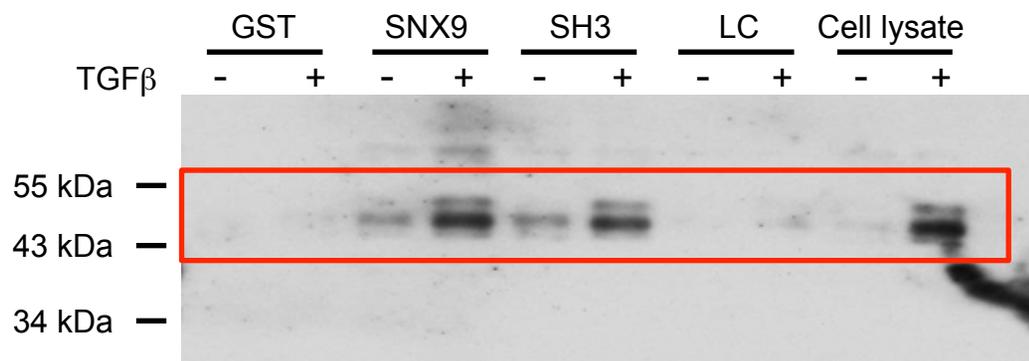


Figure S7. Effect of TAT peptides on mouse lung histopathology. (A) C57BL/6 mice were intratracheally treated with 0.9% normal saline as described in Methods. On day 14 animals were daily-administered vehicle (Methocel/saline) or Methocel/1.0 mg/kg of the indicated TAT peptide (SH3-2 or SH3-2M) intraperitoneally. Lung tissue was harvested at day 28 and subjected to Hematoxylin and Eosin (H&E) staining for histology and Masson's Trichrome (MT) for collagen. Representative images from 5 animals are shown. Scale bar: 200 μ m. (B) Hematoxylin and Eosin (H&E), Masson's Trichrome (MT), and fibronectin staining on a greater range of lung tissue from Figure 7A. Scale bar: 500 μ m.

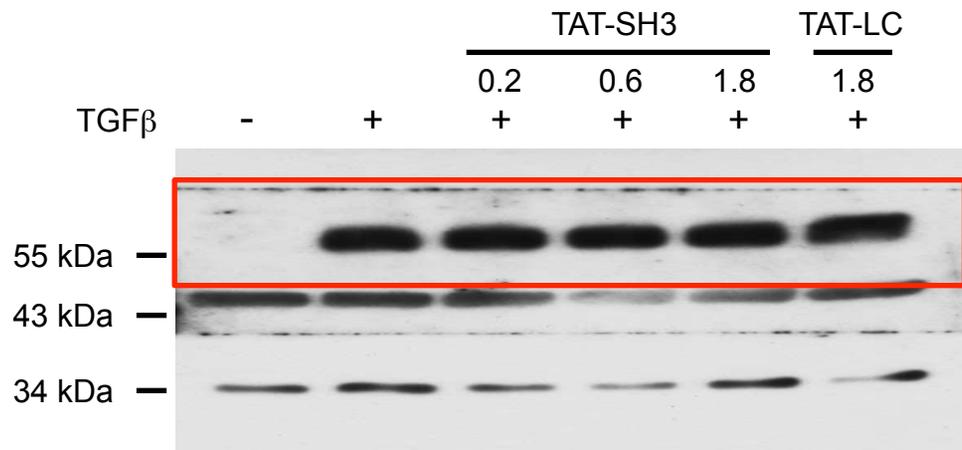


Full unedited gel for Figure 1A pSmad3

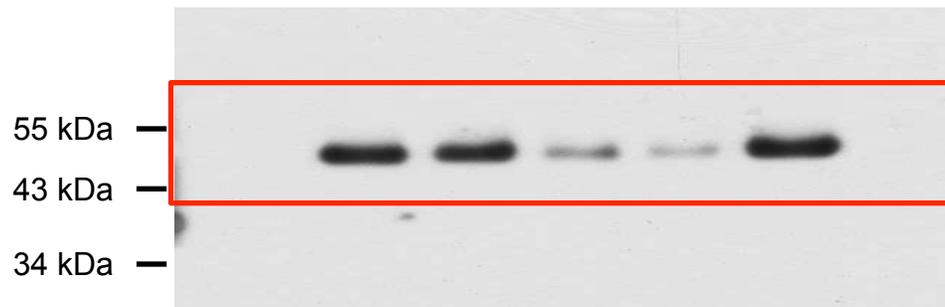


Full unedited gel for Figure 1A pSmad2

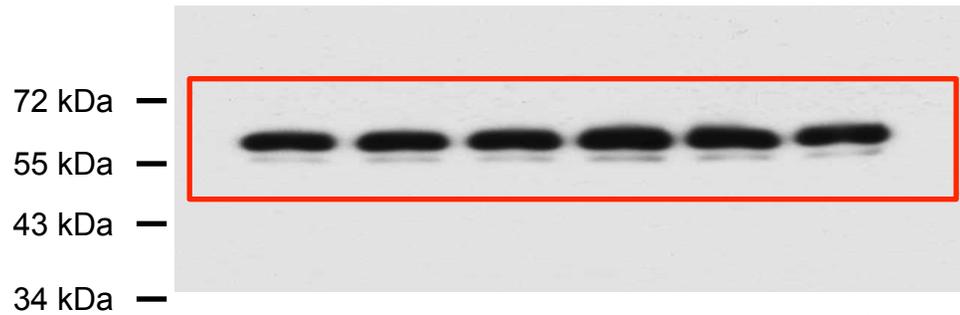
Figure 1A



Full unedited gel for Figure 1B pSmad2



Full unedited gel for Figure 1B pSmad3



Full unedited gel for Figure 1B HDAC

Figure 1B

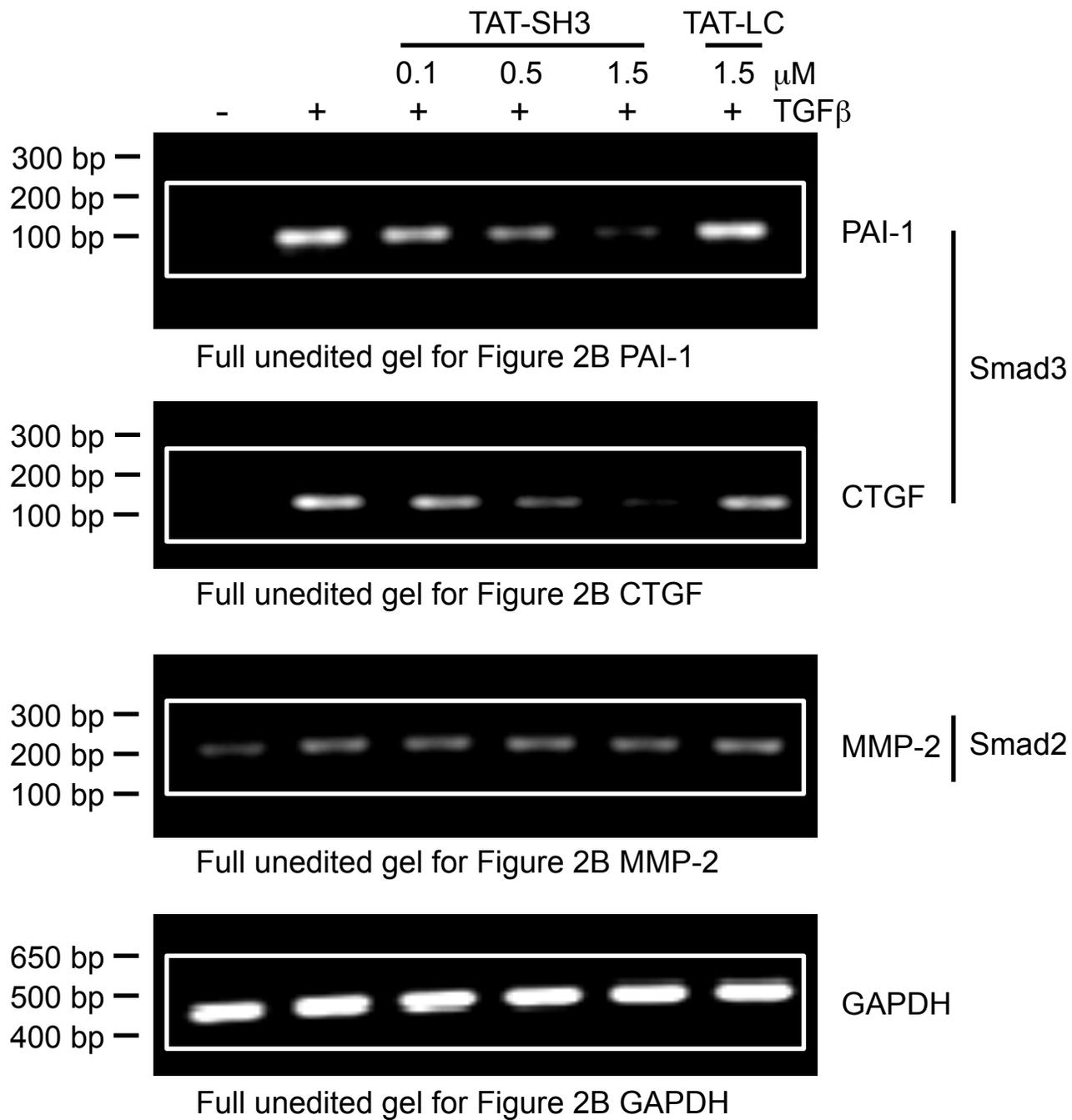
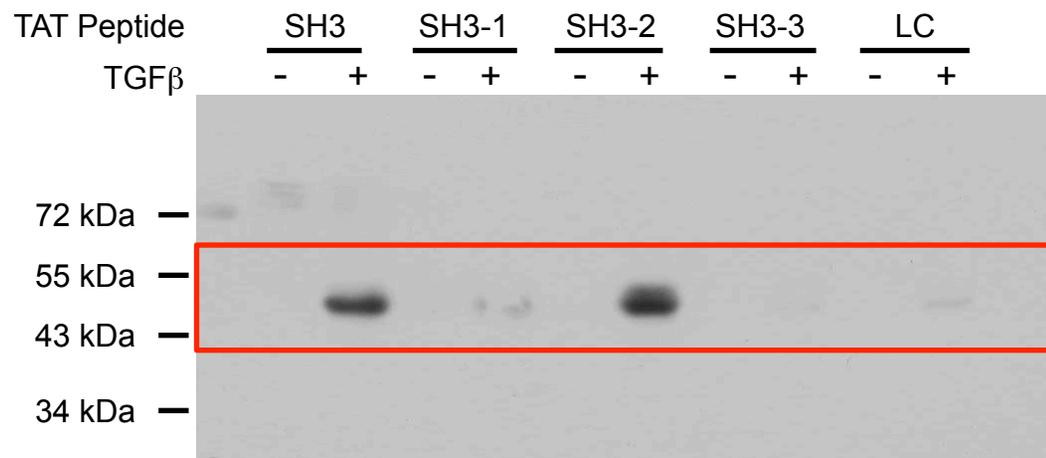
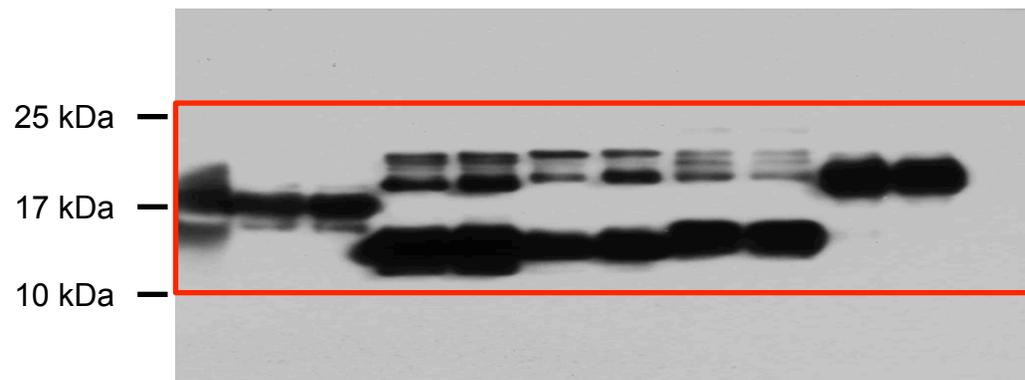


Figure 2B

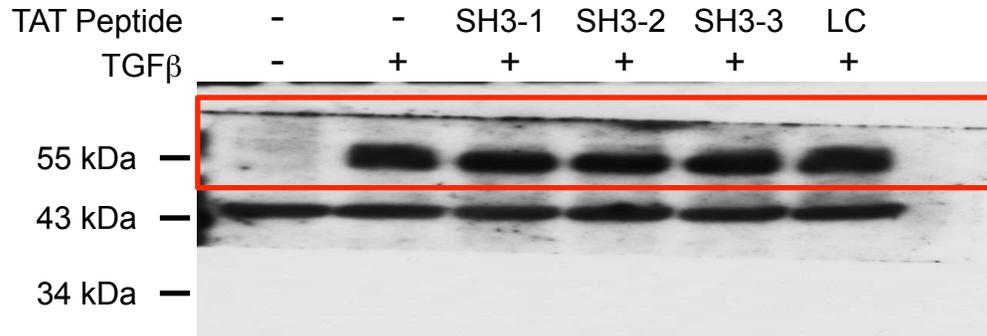


Full unedited gel for Figure 3A pSmad3

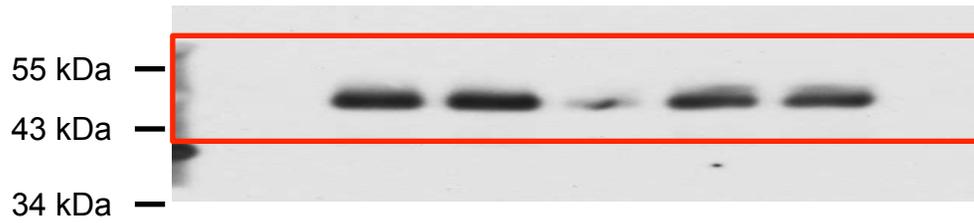


Full unedited gel for Figure 3A His

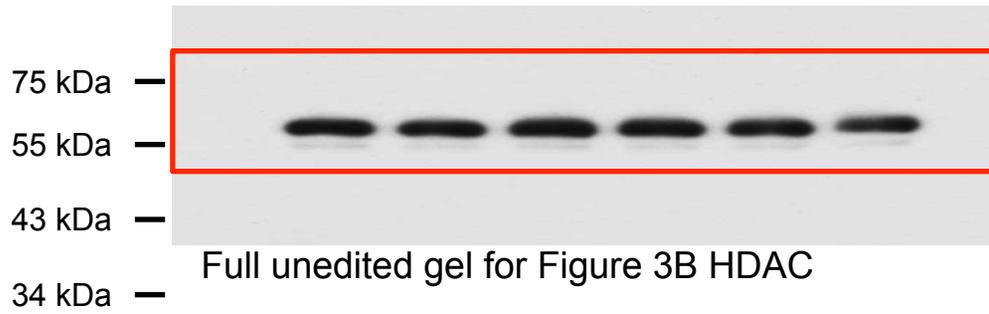
Figure 3A



Full unedited gel for Figure 3B pSmad2



Full unedited gel for Figure 3B pSmad3



Full unedited gel for Figure 3B HDAC

Figure 3B

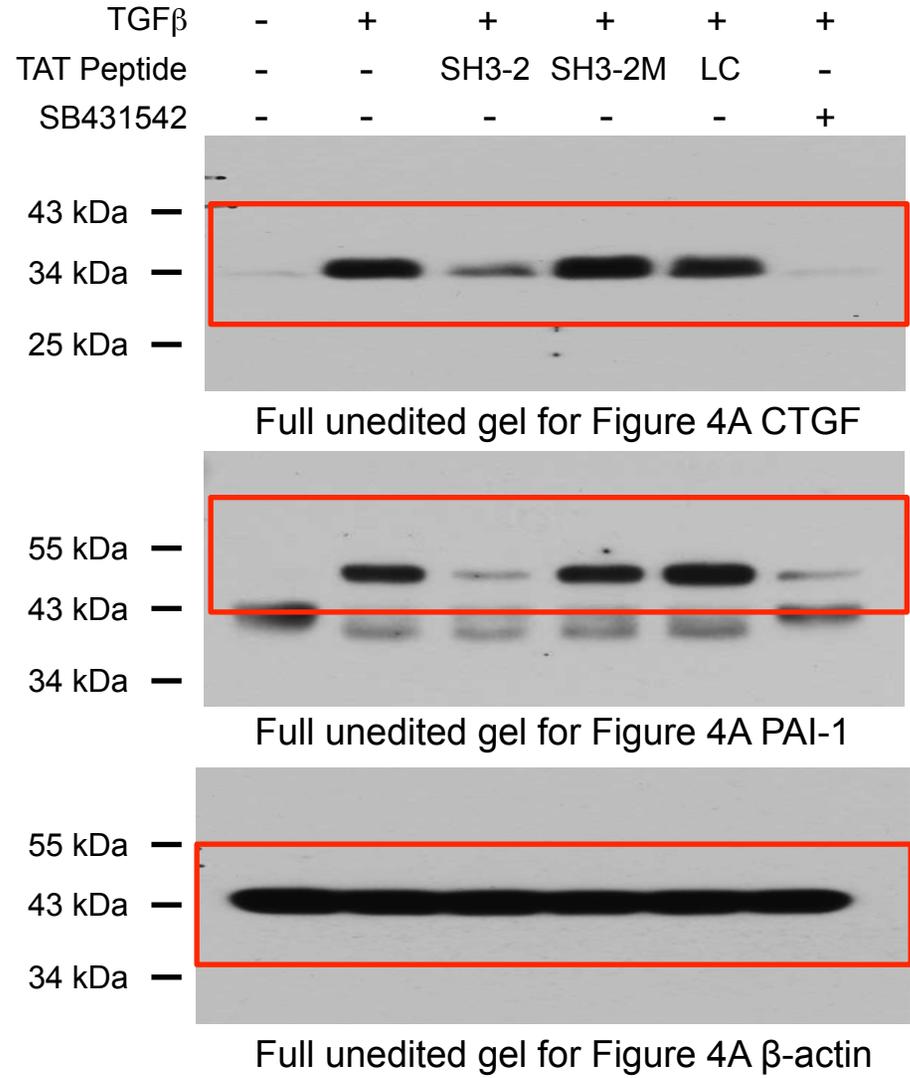
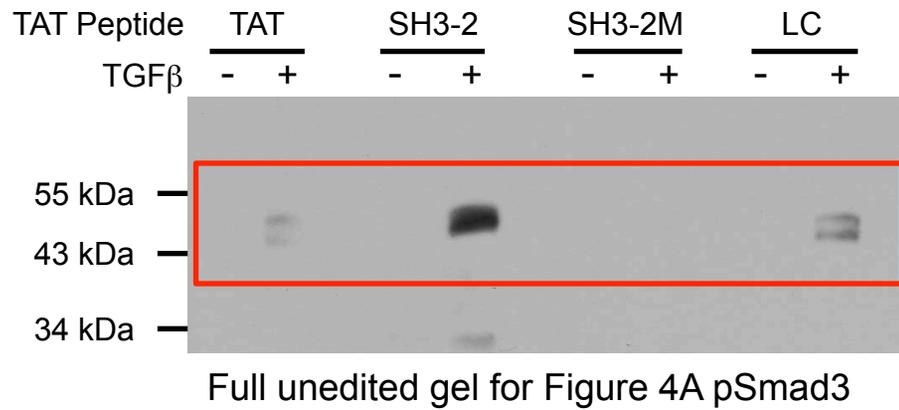


Figure 4A

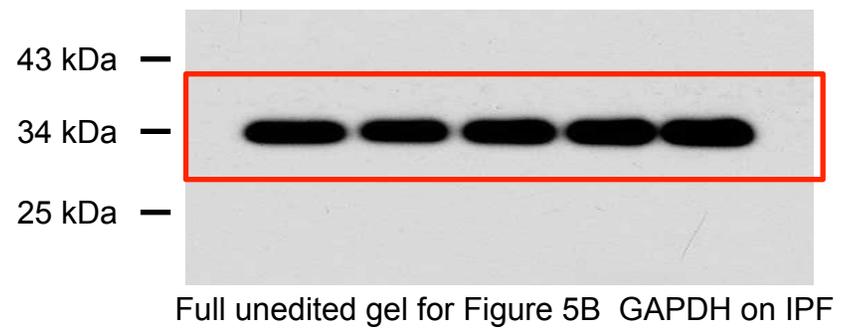
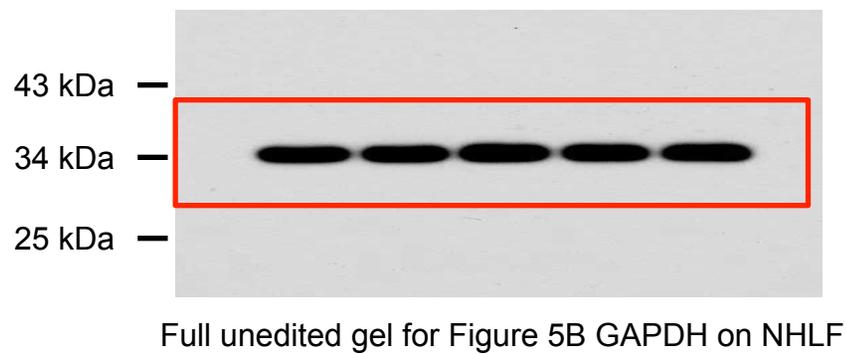
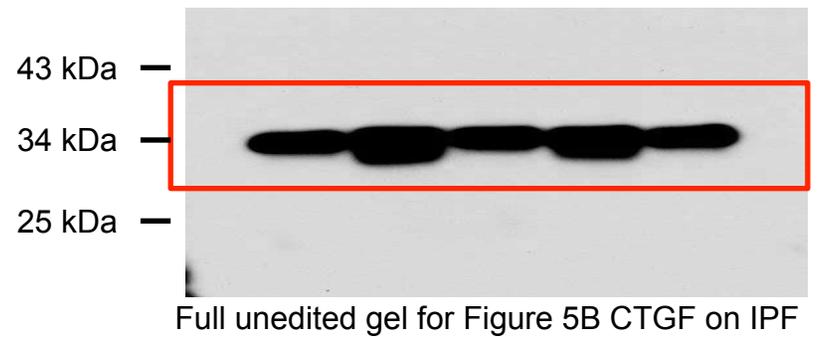
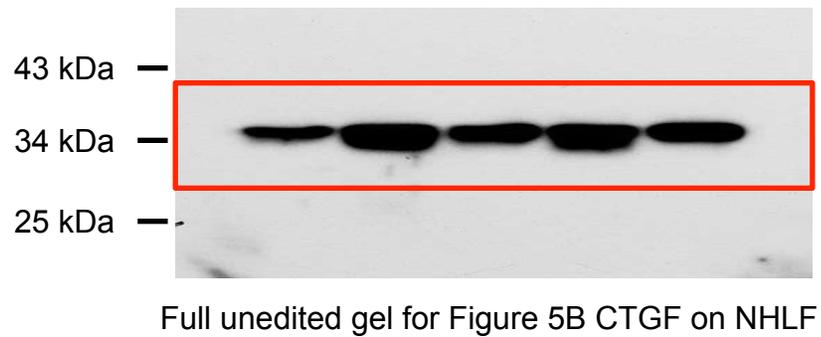
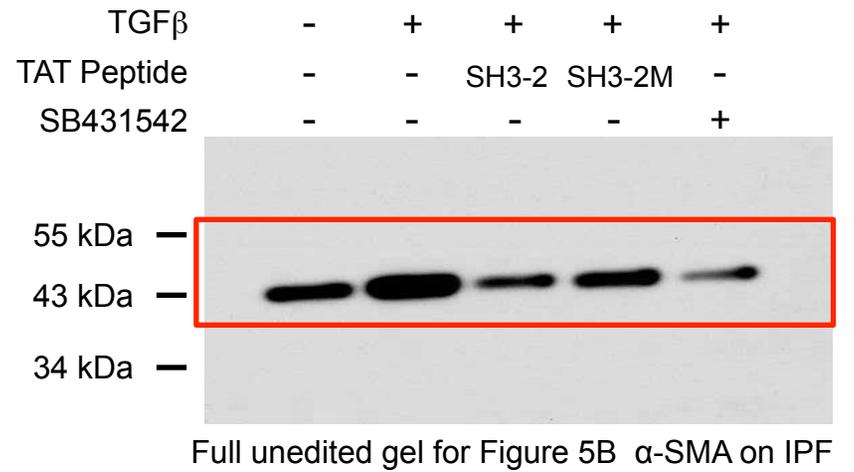
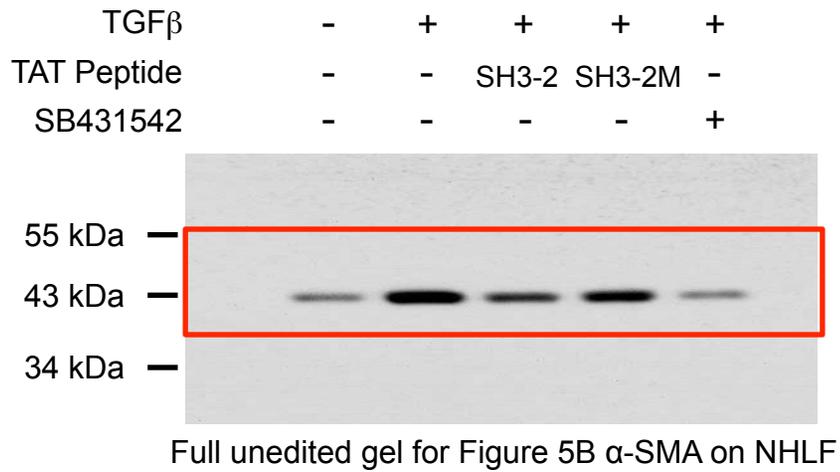


Figure 5B

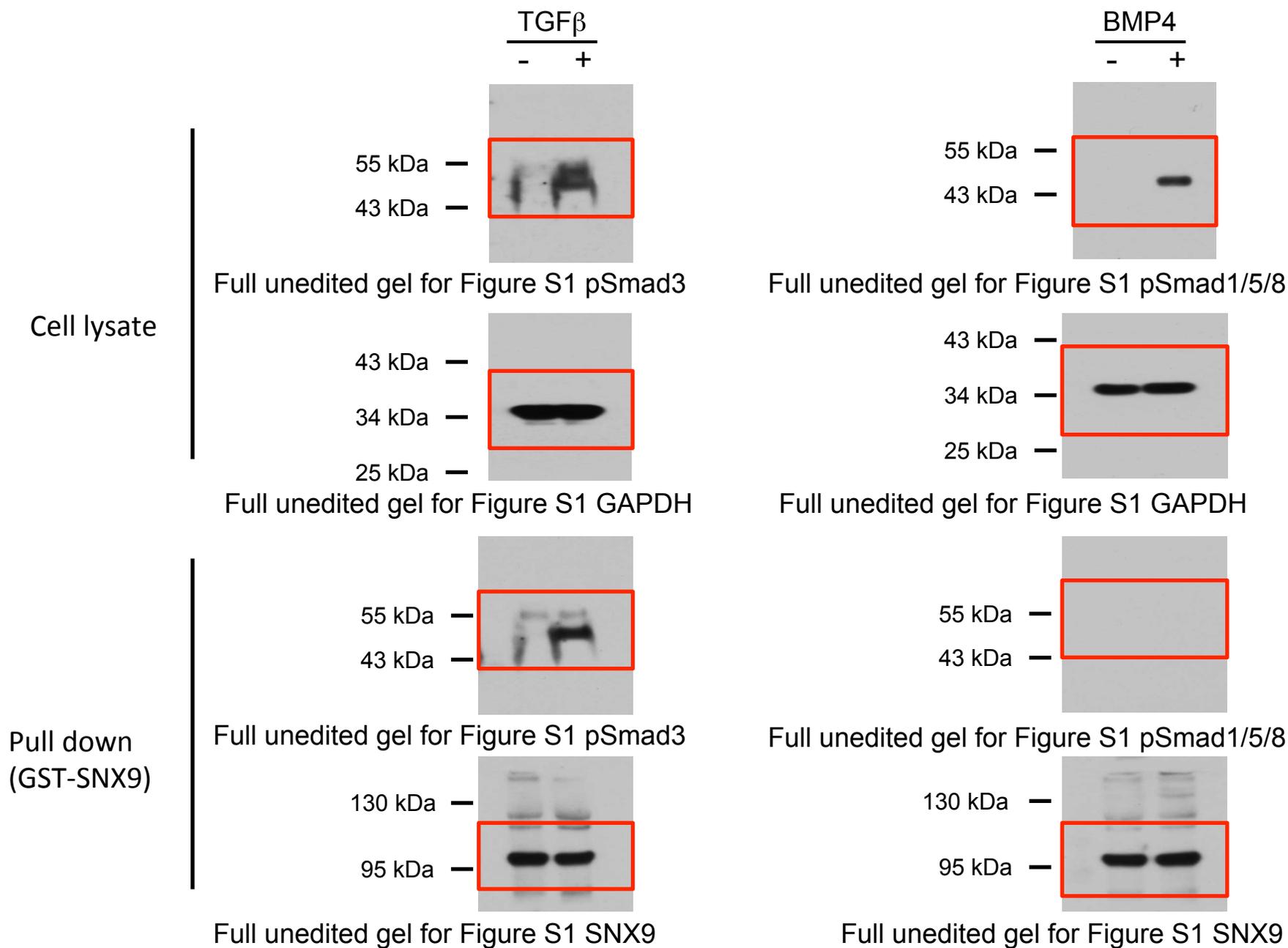
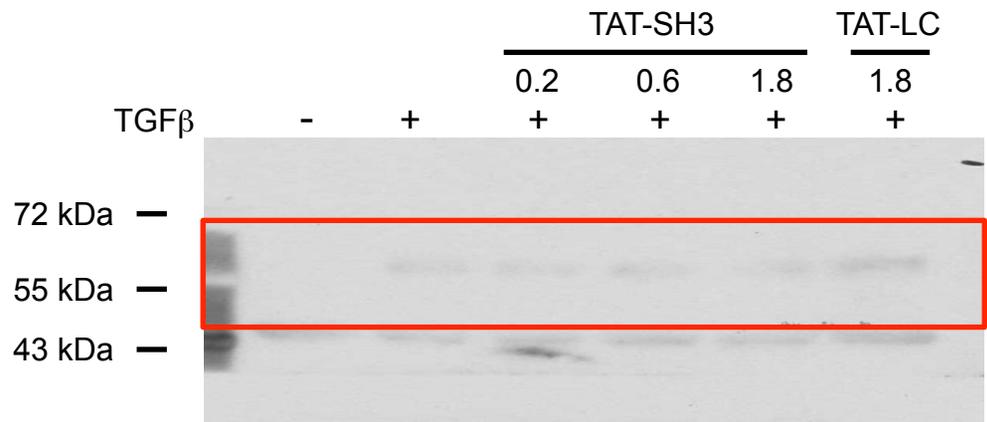
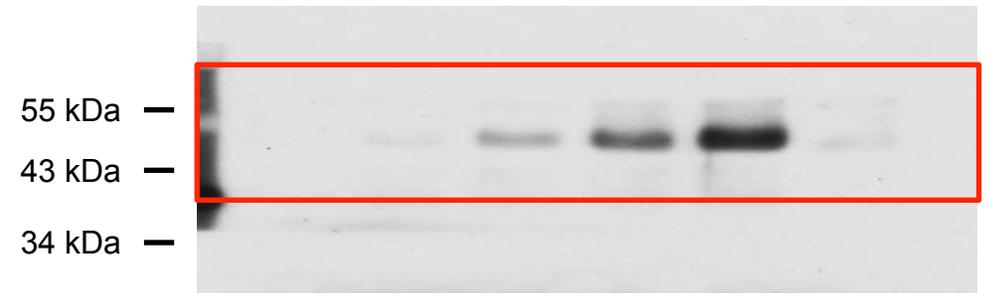


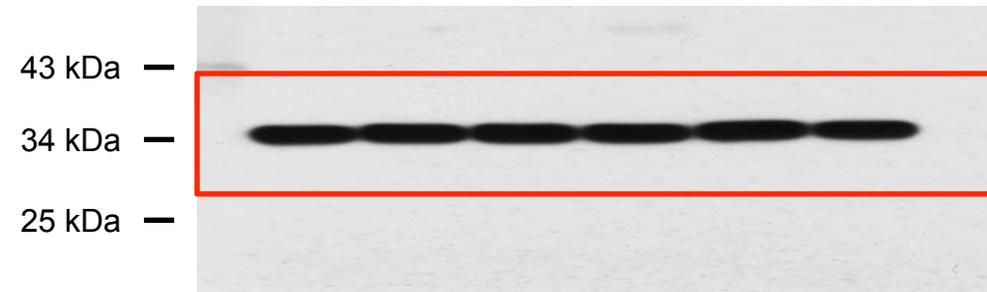
Figure S1



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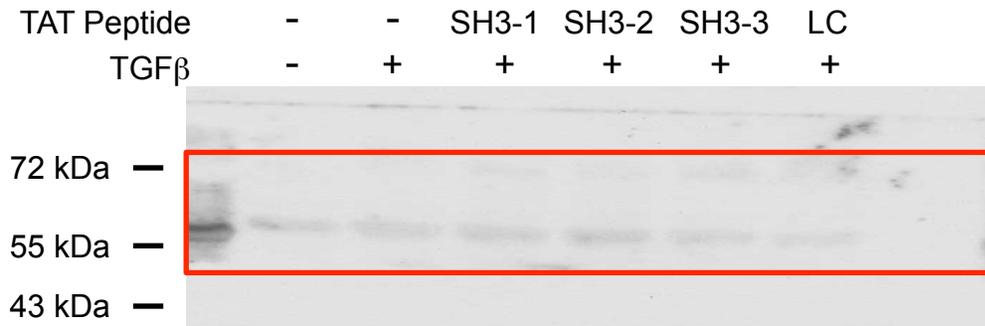


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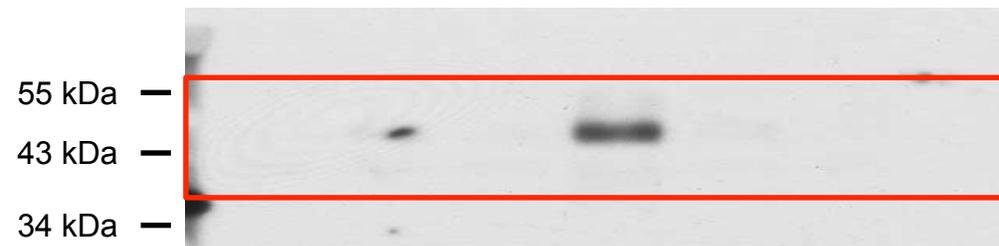


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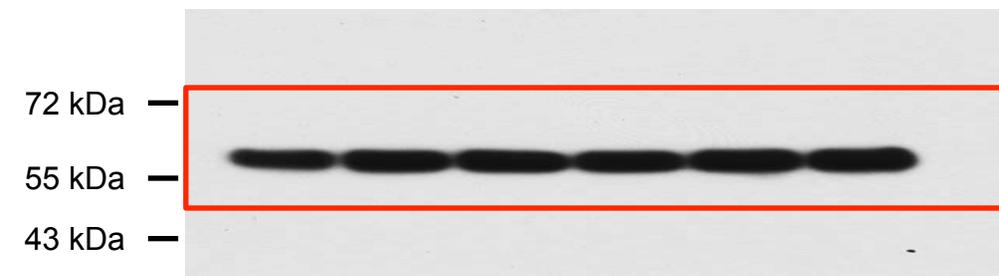
Figure S2



Full unedited gel for Figure S4 pSmad2



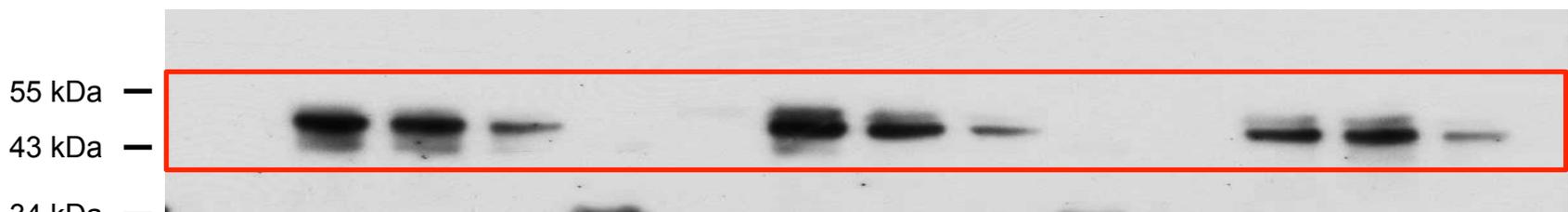
Full unedited gel for Figure S4 pSmad3



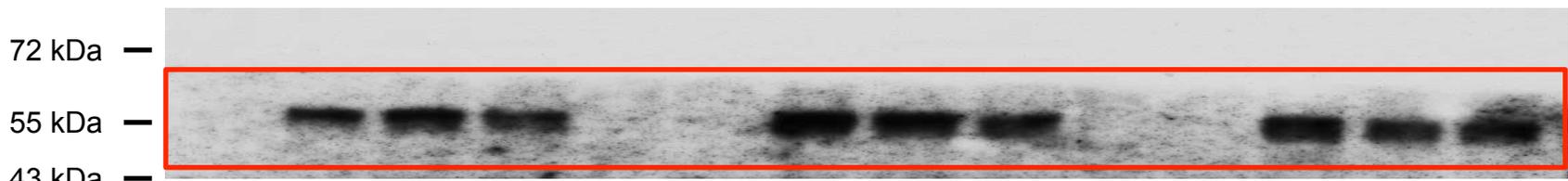
Full unedited gel for Figure S4 GAPDH

Figure S4

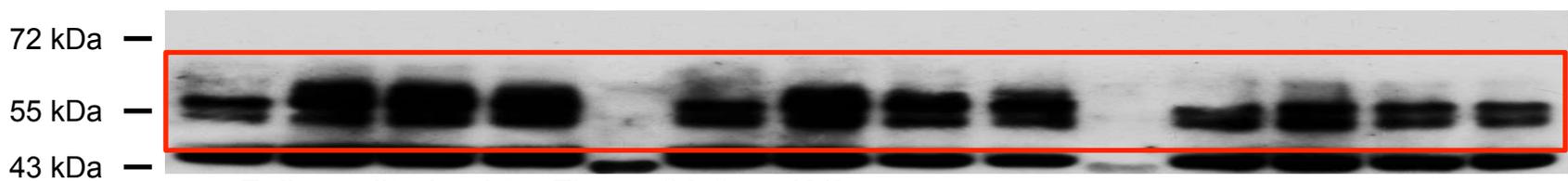
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SH3-2M	-	-	-	-	-	-	-	-	+	+	+	+



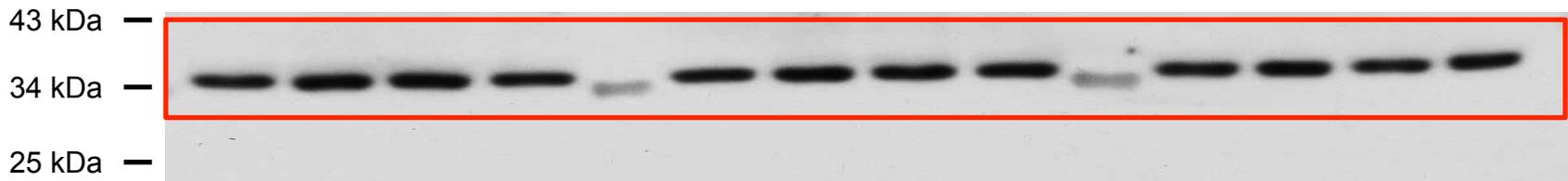
Full unedited gel for Figure S6 pSmad3



Full unedited gel for Figure S6 pSmad2



Full unedited gel for Figure S6 Smad2/3



Full unedited gel for Figure S6 GAPDH

Figure S6